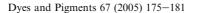


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Effect of chemical modification on dye adsorption capacity of peanut hull

Renmin Gong^{a,b,*}, Yingzhi Sun^a, Jian Chen^b, Huijun Liu^a, Chao Yang^a

^aCollege of Life Science, Anhui Normal University, Wuhu 241000, PR China ^bSchool of Biotechnology, Southern Yangtze University, Wuxi 214036, PR China

Received 27 July 2004; received in revised form 13 August 2004; accepted 9 December 2004 Available online 19 February 2005

Abstract

In this paper, the roles played by three major functional groups (amino, carboxyl and hydroxyl groups) in the biomass of peanut hull in adsorption of six dyes were investigated. These functional groups in peanut hull were chemically modified individually to determine their contribution to the adsorption of ionic dyes. The dyes used were methylene blue (MB), brilliant cresyl blue (BCB), neutral red (NR), amaranth (Am), sunset yellow (SY) and fast green FCF (FG). It was found that carboxyl group inhibited the adsorption of anionic dyes but it was major functional group in the adsorption of cationic dyes, hydroxyl group was important functional group in the adsorption of all six dyes and the effect of methylation of amino group was not significant on the adsorption of six dyes.

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Keywords: Chemical modification; Dye adsorption; Peanut hull; IR spectrum; XRD

1. Introduction

Dyes are a kind of organic compounds which can bring bright and firm color to other substances. Synthetic dyes usually have a complex aromatic molecular structure which possibly comes from coaltar based hydrocarbons such as benzene, naphthalene, anthracene, toluene, xylene, etc. The complex aromatic molecular structures of dyes make them more stable and more difficult to biodegrade [1,2]. Today there are more than 10,000 dyes available commercially [3]. Synthetic dyes have been increasingly used in the textile, leather, paper, rubber, plastics, cosmetics, pharmaceuticals and food industries. The extensive use of dyes often poses pollution problems in the form of colored wastewater

discharged into environmental water bodies. For some dyes, the dye concentration of less than 1 ppm in receiving water bodies is highly visible, so that even small quantities of dyes can color large water bodies. This not only affects aesthetic merit but also inhibits sunlight penetration and reduces photosynthetic action. In addition, some dyes or their metabolites are either toxic or mutagenic and carcinogenic [4,5].

The conventional methods for removal of dyes from wastewaters include coagulation and flocculation [6], oxidation or ozonation [7,8], membrane separation [9] and adsorption [10]. Activated carbon is popular and effective dye sorbent, but its relatively high price, high operating costs and problems with regeneration hamper its large scale application. Therefore, there is a growing need in finding low cost, renewable, locally available materials as sorbent for the removal of dye colors.

Some low cost botanic materials had directly been used as sorbent for dye adsorption from wastewater, which included apple pomace, wheat straw [11], orange

^{*} Corresponding author. College of Life Science, Anhui Normal University, Wuhu 241000, PR China. Tel.: +86 553 5016692.

E-mail address: rm.gong.nju@163.com (R. Gong).

peel [12], banana peel [13], maize cob [3], maize stalk [14], rice husk [15], barley husk [16], peanut hull [17], wood chip [18], palm fruit bunch [19,20], sawdust [21], bark [22], leaf [23], coir pith [24], banana pith [25], bagasse pith [26] and aquatic plants [27,28]. But few researches had been done about the interaction between functional groups in biomaterial and a variety of dyes. In this paper, the roles played by three major functional groups (amino, carboxyl and hydroxyl groups) in the biomass of peanut hull in adsorption of six dyes were investigated. The aim of this study was to identify the possible dye adsorption mechanisms of peanut hull. The dyes selected as sorbate were three cationic dyes: methylene blue (MB), brilliant cresyl blue (BCB), neutral red (NR) and three anionic dyes: amaranth (Am), sunset yellow (SY), fast green FCF (FG).

2. Materials and methods

2.1. Preparation of dye solutions

The dyes used in this study are listed in Table 1. Their chemical structures are shown in Fig. 1. Six dyes (MB, BCB, NR, Am, SY and FG), in commercial purity, were used without further purification. The dye stock solutions were prepared by dissolving accurately weighted dyes in distilled water to the concentration of 200 mg/l. The experimental solutions were obtained by diluting the dye stock solutions in accurate proportions to needed initial concentrations. The initial pH of each dye solution was adjusted with 0.1 M HNO₃ or NaOH using pH meter to its effective adsorption pH value obtained from the results of earlier experiments.

2.2. Preparation of peanut hull sorbent

The peanut hull used in this study was obtained from a local market. The collected biomaterial was extensively washed with tap water to remove soil and dust, sprayed with distilled water then dried in an oven at 80 °C to a constant weight. Dry biomass was crushed into powder and sieved to different particle sizes, then the biomaterial of uniform size (80–100 mesh) was preserved in the desiccator for further chemical modification.

Table 1
The general data of six dyes used in this study

Trade name	Classification	C.I. no	FW	λ_{max} (nm)
MB	Phenothiazine	52015	373.9	670
BCB	Phenoxazine	51010	317.8	630
NR	Phenazine	50040	288.8	530
Am	Monoazo	16185	604.49	520
SY	Monoazo	15985	452.37	490
FG	Triphenylmethane	42053	808.86	620

$$C_2H_5$$
 $N-CH_2$
 C_2H_5
 C_2H_5

Fig. 1. Chemical structures of the six dyes used in this study.

2.3. Chemical modification of the biomaterial

2.3.1. Methylation of amino group

The modification of amino group was made according to the same method previously reported [29,30], by shaking at ambient temperature 9 g (dry weight) of the raw biomass in 180 ml of formaldehyde (HCHO) and 360 ml of formic acid (HCOOH) for 6 h at 125 rpm. Then the treated biomaterial was thoroughly washed with distilled water, filtered and dried. This treatment resulted in methylation of amino group. The general reaction scheme is:

$$RNH2+2HCHO+2HCOOH \rightarrow RN(CH3)2 +2CO2+2H2O$$

2.3.2. Esterification of carboxyl group

The carboxyl group of the biomass was methanol esterified following a similar method previously described [31]. Esterification was carried out by heating biomass of 9 g suspended in 633 ml of 99.9% pure methanol and 5.4 ml of concentrated hydrochloric acid (HCl) given a final acidic concentration of 0.1 M HCl under reflux for 48 h. Then the esterified biomaterial was thoroughly washed with distilled water, filtered and dried. The general reaction scheme of this treatment is:

$$RCOOH + CH_3OH \xrightarrow{H+} RCOOCH_3 + H_2O$$

2.3.3. Acetylation of amino and hydroxyl group

According to the method previously used [32], total acetylation of amino and hydroxyl group of the sorbent was carried out by refluxing the biomass suspension in acetic anhydride at 80 °C for 10 h. Then the acetylated biomaterial was thoroughly washed with distilled water, filtered and dried. The general reaction scheme of this treatment is:

$$RNH_2 + (CH_3CO)_2O \rightarrow RNHCOCH_3 + CH_3COOH$$

 $RCH_2OH + (CH_3CO)_2O \rightarrow RCH_2OCOCH_3$
 $+ CH_3COOH$

2.4. Dye adsorption experiments

The adsorption experiments of cationic dyes were carried out in a rotary shaker at 150 rpm and at ambient temperature using 250 ml shaking flasks containing 100 ml dye solutions. The concentration and initial pH value of dye solution were 100 mg/l and 5.0, respectively. Four flasks were used for each dye. Raw, methylated, esterified and acetylated biomaterials (0.2 g) were respectively added to four flasks of each dye, then the flasks were sealed up to prevent change of volume of the solution during the experiments. After shaking the flasks for 24 h, the samples were withdrawn from the flasks and the dye solutions were separated from the sorbent by

filtration with a 200 mesh/inch stainless steel sieve then centrifugation. Dye concentrations in the supernatant solutions were determined.

The adsorption experiments of anionic dyes were carried out following a similar method as cationic dye adsorption, but the concentration and initial pH value of dye solution were 50 mg/l and 2.0, respectively. Furthermore, 0.5 g of raw, methylaed, esterified and acetylated biomaterials were used and the adsorption time was 36 h.

2.5. Dye concentration determination

Dye concentrations were estimated by measuring adsorbance at maximum wavelengths of dyes with a 752W UV—vis Grating Spectrophotometer (Shanghai, China) and computing from the calibration curves. The amounts of dyes sorbed by the biomaterials were calculated using the following equation:

$$q = (C_0 - C_e)V/W$$

where q (mg/g) is the amount of dye sorbed by biomass, C_0 and C_e (mg/l) are the initial and equilibrium liquidphase concentration of dye, respectively, V (l) is the initial volume of dye solution, and W (g) is the weight of the biomass.

The experiments were conducted in duplicate and the negative controls (with no sorbent) and were simultaneously carried out to ensure that adsorption was by peanut hull biomass and not by the container.

2.6. IR spectra and XRD study

The IR spectra of raw and chemically modified sorbents were obtained using a Fourier transform infrared spectrometer (BIO-RAD FTS-40). For IR spectra, 5 mg of biomass was encapsulated in 400 mg of KBr. Translucent disk was made by pressing the

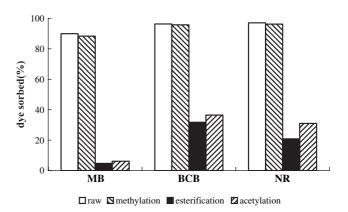


Fig. 2. Effect of chemical modification on adsorption of MB, BCB, NR by peanut hull.

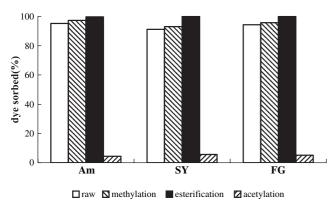


Fig. 3. Influence of chemical modification on adsorption of Am, SY, FG by peanut hull.

ground mixed material with the aid of a bench press (955 kg for 10 min).

Crystallinities of the raw and chemically modified sorbents were determined by X-ray diffraction using a diffractometer (X'TRA ARL) operated at 50 kV and 40 mA. The scanning scope and scanning speed were 5–55° and 10°/min, respectively, using Cu K_{\alpha} radiation.

3. Results and discussion

3.1. Effect of dye adsorption by chemical modification

The effects of chemical modification on the removal ratios of cationic dyes are shown Fig. 2. None of chemical modification increased dye adsorption ratios. After amino group methylation, removal capacities of three dyes decreased, but only a little. Esterification of carboxyl group induced rapid decrease of three dye adsorptions; this experimental result proved that carboxyl group was major functional group in the adsorption of cationic dyes. Total acetylation of amino and hydroxyl groups also decreased the adsorption percentages of all three dyes, but decrease extent was less than that of esterification, it indicated that hydroxyl group also was important functional group in the adsorption of cationic dyes.

Fig. 3 showed the influences of chemical modification on adsorption percentages of anionic dyes. Methylation of amino group increased slightly the adsorption ratios of three dyes. After carboxyl group esterification, three dyes were completely removed from solution, it indicated that the carboxyl group bearing negative charge inhibited the adsorption of anionic dyes and in case removing the negative charge of carboxyl group by esterification, dye uptake capacities were obviously increased. Total acetylation of amino and hydroxyl groups extremely decreased the adsorption ratios of all three dyes, it showed that hydroxyl group was important functional group in the adsorption of anionic dyes.

3.2. Influence of IR spectra and XRD by chemical modification

The IR spectra of raw and chemically modified biomaterials are shown in Figs. 4–7. From Fig. 5, it could been seen, the broad mixed stretching vibration adsorption band of amino and hydroxyl groups at 3392 cm⁻¹ was reduced, it was the result of methylation. Fig. 6 showed that esterification brought reduction of stretching vibration adsorption band of carboxyl group at 1730 cm⁻¹. Fig. 7 indicated that the stretching

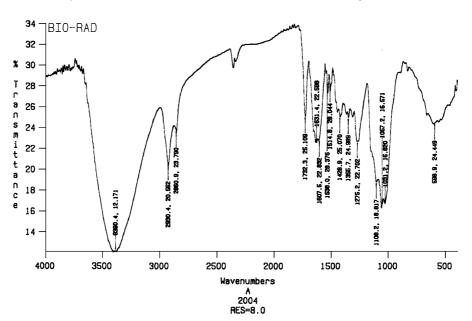


Fig. 4. The IR spectrum of raw sorbent.

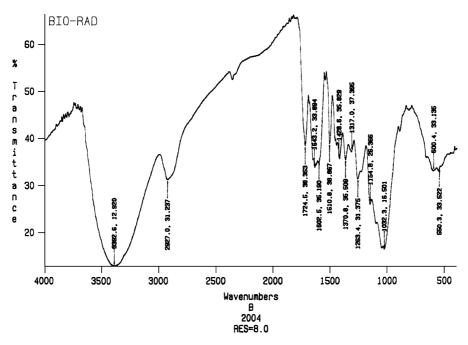


Fig. 5. The IR spectrum of methylation sorbent.

vibration adsorption band of carboxyl group at $1746~\rm cm^{-1}$ was obviously increased due to acetylation of amino and hydroxyl groups, and besides, an increase of the adsorption band at $1235~\rm cm^{-1}$ also could been found.

The XRD diagrams of raw and chemically modified sorbents are shown in Fig. 8. The XRD pattern of raw sorbent showed typical spectrum of cellulosic material, having main and secondary peaks at 2θ of 22° and 16° , respectively [33]. The main peak is taken as indicative of

the presence of highly organized crystalline cellulose, while the secondary peak is a measure of a less organized polysaccharide structure. The XRD diagram of methylation sorbent was very similar to that of raw sorbent. After carboxyl group esterification, the main and secondary peak heights in XRD diagram was increased, it indicated that the crystallinity of esterified sorbent was actually increased. Total acetylation of amino and hydroxyl groups induced disappearance of the secondary peak in XRD diagram.

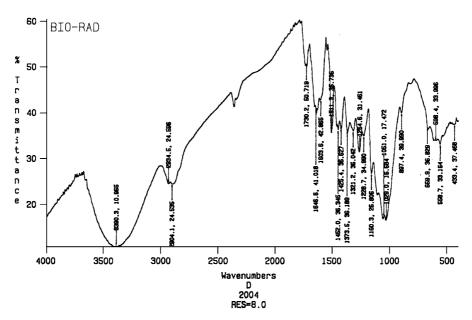


Fig. 6. The IR spectrum of esterification sorbent.

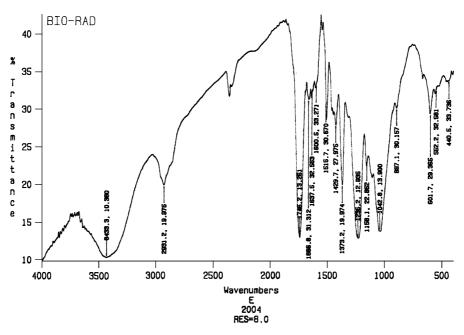


Fig. 7. The IR spectrum of acetylation sorbent.

4. Conclusions

Studies on effects of chemical modification on dye adsorption on sorbent derived from peanut hull suggested the following conclusions:

• The effect of methylation of amino group on ionic dye adsorption was not significant. The possible reason was that at ambient temperature, the methylation could not be carried out completely due to the tough cell wall of peanut hull.

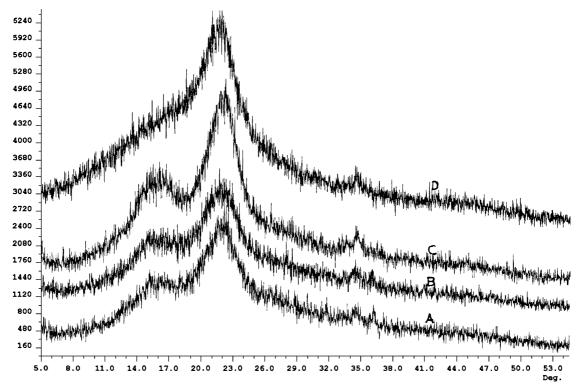


Fig. 8. XRD diagrams of raw and chemically modified sorbents. A: raw sorbent, B: methylation sorbent, C: esterification sorbent, D: acetylation sorbent.

- The carboxyl group inhibited the adsorption of anionic dyes because of its negative charge and in case removing the negative charge of carboxyl group by esterification, anionic dye uptake capacity was obviously increased. The carboxyl group was major functional group in the adsorption of cationic dyes.
- The hydroxyl group was important functional group in the adsorption of cationic and anionic dyes.

Acknowledgments

This work was financially supported by the Key Laboratory of Bioresource Protection and Utilization of Anhui Province, the Anhui Educational Bureau and the Anhui Normal University, China.

References

- Fewson CA. Biodegradation of xenobiotic and other persistent compounds: the causes of recalcitrance. Trends Biotechnol 1988;6:148-53.
- [2] Seshadri S, Bishop PL, Agha AM. Anaerobic/aerobic treatment of selected azo dyes in wastewater. Waste Manage 1994;15:127–37.
- [3] Nigam P, Armour G, Banat IM, Singh D, Marchant R. Physical removal of textile dyes from effluents and solid-state fermentation of dye-adsorbed agricultural residues. Bioresour Technol 2000;72:219–26.
- [4] Chen KC, Wu JY, Huang CC, Liang YM, Hwang SCJ. Decolorization of azo dye using PVA-immobilized microorganisms. J Biotechnol 2003;101:241–52.
- [5] Heiss GS, Gowan B, Dabbs ER. Cloning of DNA from a *Rhodococcus* strain conferring the ability to decolorize sulfonated azo dyes. FEMS Microbiol Lett 1992;99:221–6.
- [6] Panswed J, Wongchaisuwan S. Mechanism of dye wastewater color removal by magnesium carbonate-hydrated basic. Water Sci Technol 1986;18:139—44.
- [7] Malik PK, Saha SK. Oxidation of direct dyes with hydrogen peroxide using ferrous ion as catalyst. Sep Purif Technol 2003;31:241-50.
- [8] Koch M, Yediler A, Lienert D, Insel G, Kettrup A. Ozonation of hydrolyzed azo dye reactive yellow 84 (CI). Chemosphere 2002;46:109–13.
- [9] Ciardelli G, Corsi L, Marucci M. Membrane separation for wastewater reuse in the textile industry. Resour Conserv Recycl 2000;31:189-97.
- [10] Venkata RB, Sastray CA. Removal of dyes from water and wastewater by adsorption. Indian J Environ Prot 1987;7:363-76.
- [11] Robinson T, Chandran B, Nigam P. Removal of dyes from a synthetic textile dye effluent by biosorption on apple pomace and wheat straw. Water Res 2002;36:2824-30.
- [12] Namasivayam C, Muniasamy N, Gayatri K, Rani M, Ranganathan K. Removal of dyes from aqueous solutions by cellulosic waste orange peel. Bioresour Technol 1996;57:37–43.

- [13] Annadurai G, Juang R, Lee D. Use of cellulose-based wastes for adsorption of dyes from aqueous solutions. J Hazard Mater B 2002:92:263-74.
- [14] Meyer V, Carlsson FHH, Oellermann RA. Decolourization of textile effluent using a low cost natural adsorbent material. Water Sci Technol 1992;26:1205–11.
- [15] McKay G, Ramprasad G, Mowli P. Desorption and regeneration of dye colours from low-cost materials. Water Res 1987;21: 375-7.
- [16] Robinson T, Chandran B, Nigam P. Effect of pretreatments of three waste residues, wheat straw, corncobs and barley husks on dye adsorption. Bioresour Technol 2002;85:119–24.
- [17] Gong RM, Ding Y, Li M, Yang C, Liu HJ, Sun YZ. Utilization of powdered peanut hull as biosorbent for removal of anionic dyes from aqueous solution. Dyes Pigments 2005;64:187–92.
- [18] Poots VJP, McKay G, Healy JJ. The removal of acid dye from effluent using natural adsorbents — II wood. Water Res 1976;10:1067—70.
- [19] Nassar MM, Hamoda MF, Radwan GH. Adsorption equilibria of basic dyestuff onto palm-fruit bunch particles. Water Sci Technol 1995;32:27–32.
- [20] Nassar MM. Intraparticle diffusion of basic red and basic yellow dyes on palm fruit bunch. Water Sci Technol 1999;40:133–9.
- [21] Garg VK, Gupta R, Yadav AB, Kumar R. Dye removal from aqueous solution by adsorption on treated sawdust. Bioresour Technol 2003;89:121–4.
- [22] Morais LC, Freitas OM, Goncalves EP, Vasconcelos LT, Beca CGG. Reactive dyes removal from wastewaters by adsorption on eucalyptus bark: variables that define the process. Water Res 1999;33:979–88.
- [23] Bhattacharyya KG, Sarma A. Adsorption characteristics of the dye, brilliant green, on neem leaf powder. Dyes Pigments 2003;57:211–22.
- [24] Namasivayam C, Kadirvelu K. Coir pith, an agricultural waste by-product, for the treatment of dyeing wastewater. Bioresour Technol 1994;48:79–81.
- [25] Namasivayam C, Prabha D, Kumutha M. Removal of direct red and acid brilliant blue by adsorption on to banana pith. Bioresour Technol 1998;64:77–9.
- [26] McKay G, El Geundi E, Nassar MM. Pore diffusion during the adsorption of dyes onto bagasse pith. Process Saf Environ 1996;74:487–502.
- [27] Low KS, Lee CK, Heng LL. Sorption of basic dyes by *Hydrilla* verticillata. Environ Technol 1993;14:115–24.
- [28] Low KS, Lee CK, Tan KK. Biosorption of basic dye by water hyacinth roots. Bioresour Technol 1995;52:79–83.
- [29] Kapoor A, Viraraghavan T. Heavy metal biosorption sites in *Aspergillus niger*. Bioresour Technol 1997;61:221–7.
- [30] Fu Y, Viraraghavan T. Dye biosorption sites in Aspergillus niger. Bioresour Technol 2002;82:139–45.
- [31] Tiemann KJ, Gamez G, Dokken K, Parsons JG, Gardea-Torresdey JL. Chemical modification and X-ray absorption studies for lead(II) binding by *Medicago sativa* (alfalfa) biomass. Microchem J 2002;71:287–93.
- [32] Bai RS, Abraham TE. Studies on enchancement Cr(VI) biosorption by chemically modified biomass of *Rhizopus nigricans*. Water Res 2002;36:1224–36.
- [33] Atalla RH. The structure of cellulose: recent development. In: Soltes EJ, editor. Wood and agricultural residues — research on use for feed, fuels, and chemicals. New York: Academic; 1983. p. 59-77.